

Online Research @ Cardiff

This is an Open Access document downloaded from ORCA, Cardiff University's institutional repository: <https://orca.cardiff.ac.uk/id/eprint/79478/>

This is the author's version of a work that was submitted to / accepted for publication.

Citation for final published version:

García-Ramón, Diana C., Palma, Leopoldo, Berry, Colin ORCID: <https://orcid.org/0000-0002-9943-548X>, Osuna, Antonio and Vílchez, Susana 2015. Draft genome sequence of the entomopathogenic bacterium *Bacillus pumilus* 15.1, a strain highly toxic to the mediterranean fruit fly *Ceratitis capitata*. *Genome Announcements* 3 (5) , -15. 10.1128/genomeA.01019-15 file

Publishers page: <http://dx.doi.org/10.1128/genomeA.01019-15>
<<http://dx.doi.org/10.1128/genomeA.01019-15>>

Please note:

Changes made as a result of publishing processes such as copy-editing, formatting and page numbers may not be reflected in this version. For the definitive version of this publication, please refer to the published source. You are advised to consult the publisher's version if you wish to cite this paper.

This version is being made available in accordance with publisher policies.

See

<http://orca.cf.ac.uk/policies.html> for usage policies. Copyright and moral rights for publications made available in ORCA are retained by the copyright holders.



Draft Genome Sequence of the Entomopathogenic Bacterium *Bacillus pumilus* 15.1, a Strain Highly Toxic to the Mediterranean Fruit Fly *Ceratitis capitata*

Diana C. García-Ramón,^a Leopoldo Palma,^{b,c} Colin Berry,^d Antonio Osuna,^a Susana Vilchez^{a,e}

Institute of Biotechnology, Campus Fuentenueva, University of Granada, Granada, Spain^a; Facultad de Ciencias Agrarias, Universidad Nacional del Litoral, Esperanza, Santa Fe, Argentina^b; Consejo Nacional de Investigaciones Científicas y Técnicas, Buenos Aires, Argentina^c; Cardiff School of Biosciences, Cardiff University, Cardiff, Wales, United Kingdom^d; Department of Biochemistry and Molecular Biology I, Campus Fuentenueva, University of Granada, Granada, Spain^e

We present the draft whole-genome sequence of the entomopathogenic *Bacillus pumilus* 15.1 strain that consists of 3,795,691 bp and 3,776 predicted protein-coding genes. This genome sequence provides the basis for understanding the potential mechanism behind the toxicity and virulence of *B. pumilus* 15.1 against the Mediterranean fruit fly.

Received 27 July 2015 Accepted 28 July 2015 Published 24 September 2015

Citation García-Ramón DC, Palma L, Berry C, Osuna A, Vilchez S. 2015. Draft genome sequence of the entomopathogenic bacterium *Bacillus pumilus* 15.1, a strain highly toxic to the Mediterranean fruit fly *Ceratitis capitata*. *Genome Announc* 3(5):e01019-15. doi:10.1128/genomeA.01019-15.

Copyright © 2015 García-Ramón et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Susana Vilchez, svt@ugr.es.

Bacillus pumilus strain 15.1 has been recently described to be toxic against larvae of the Mediterranean fruit fly, *Ceratitis capitata*, one of the most damaging pests for fruits and vegetables worldwide (1). Strain 15.1 was isolated from a partially decomposed reed plant and has recently been characterized microbiologically and biochemically (D. C. Garcia-Ramon, C. A. Molina, A. Osuna, S. Vilchez, submitted for publication). *B. pumilus* strain 15.1 was deposited in the Spanish Type Culture Collection as CECT 7462.

B. pumilus strains are known to have very interesting properties, such as being highly resistant to environmental stresses (2) and producing a wide range of metabolites of industrial value. Nevertheless, only a few genomes from this species have been published. We report the first genome sequence of an entomopathogenic *B. pumilus* strain.

DNA from *B. pumilus* 15.1 was sequenced on a HiSeq 2000 sequencing system (Illumina Sequencing) in a single-read mode with a read length of 50 bases (GATC Biotech, Constance, Germany). The whole-genome sequencing yielded 26,322,535 reads. The reads were assembled in contigs by using Velvet software (3) with the *de novo* assembly tool and default parameters and then by iterative mapping using Geneious Pro R8 software, obtaining 63 contigs. The genome size of *B. pumilus* was 3,795,691, with a G+C content of 41.3%. Genome annotation was added by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP), although it was also analyzed with BLAST (4) using a custom insecticidal toxin database (5, 6). A total of 3,776 protein-coding genes, 45 pseudogenes, 2 rRNA genes, 16 tRNA genes, and 1 noncoding RNA gene were predicted in the *B. pumilus* 15.1 genome.

B. pumilus 15.1 bears at least two extrachromosomal elements, one plasmid and one megaplasmid (D. Garcia-Ramon, M. J. Luque-Navas, C. A. Molina, C. del Val, A. Osuna, and S. Vilchez, submitted for publication). The plasmid sequence (7,785 bp, 35.7% G+C content) corresponds to contig 38 in the assembly. The megaplasmid has not yet been assigned to any contig or contig

sequences, but it has been detected in total DNA extractions on agarose gels (D. Garcia-Ramon et al., submitted). The *B. pumilus* 15.1 genome also contains prophage elements, even though a genome analysis revealed the presence of a clustered regularly interspaced short palindromic repeat (CRISPR)/Cas system (bacterial innate immune mechanism for protection from foreign DNA) (7). Interestingly, during sporulation this strain forms parasporal crystals that morphologically resemble those produced by *Bacillus thuringiensis* Cry proteins (D. C. Garcia-Ramon, C. A. Molina, A. Osuna, S. Vilchez, submitted for publication). The role of these crystalline structures has not yet been elucidated, and we have detected no cry-like gene coding sequences in the genome. However, the *B. pumilus* 15.1 genome harbors other genes encoding well-known entomopathogenic factors, such as chitinases (8), metalloproteases (9), and cytolysins (10).

The analysis and characterization of the *B. pumilus* 15.1 genome will provide new insights to allow the elucidation of the pathogenic mechanism of *B. pumilus* 15.1 against the Mediterranean fruit fly, providing useful information for the development of novel biotechnological products suitable for the control of this pest and the discovery of unknown virulence factors of this entomopathogenic bacterium.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [LBDK000000000](https://www.ncbi.nlm.nih.gov/nuclseq/LBDK000000000/). The version described in this paper is version LBDK000000000.1.

ACKNOWLEDGMENTS

This research was supported by the Consejería de Ciencia e Innovación from Junta de Andalucía through their research Programme “Proyectos de Excelencia” (grant ID: AGR-6409).

We thank Steffan Adams (Cardiff University, United Kingdom) for assisting with remote access to the Assembler server to perform bioinformatic analyses.

REFERENCES

1. Molina CA, Caña-Roca JF, Osuna A, Vilchez S. 2010. Selection of a *Bacillus pumilus* strain highly active against *ceratitis capitata* (Wiedemann) larvae. *Appl Environ Microbiol* 76:1320–1327. <http://dx.doi.org/10.1128/AEM.01624-09>.
2. Gioia J, Yerrapragada S, Qin X, Jiang H, Igboeli OC, Muzny D, Dugan-Rocha S, Ding Y, Hawes A, Liu W, Perez L, Kovar C, Dinh H, Lee S, Nazareth L, Blyth P, Holder M, Buhay C, Tirumalai MR, Liu Y, Dasgupta I, Bokhetache L, Fujita M, Karouia F, Eswara Moorthy P, Siefert J, Uzman A, Buzumbo P, Verma A, Zwiya H, McWilliams BD, Olowu A, Clinkenbeard KD, Newcombe D, Golebiewski L, Petrosino JF, Nicholson WL, Fox GE, Venkateswaran K, Highlander SK, Weinstock GM. 2007. Paradoxical DNA repair and peroxide resistance gene conservation in *Bacillus pumilus* SAFR-032. *PLoS One* 2:e928. <http://dx.doi.org/10.1371/journal.pone.0000928>.
3. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
4. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [http://dx.doi.org/10.1016/S0022-2836\(05\)80360-2](http://dx.doi.org/10.1016/S0022-2836(05)80360-2).
5. Palma L, Muñoz D, Berry C, Murillo J, Caballero P. 2014. Draft genome sequences of two *Bacillus thuringiensis* strains and characterization of a putative 41.9-kDa insecticidal toxin. *Toxins* 6:1490–1504. <http://dx.doi.org/10.3390/toxins6051490>.
6. Palma L, Muñoz D, Murillo J, Caballero P. 2014. Draft genome sequence of *Bacillus thuringiensis* serovar Tolworthi strain Na205-3, an isolate toxic for *Helicoverpa armigera*. *Genome Announc* 2(2):e00187-14. <http://dx.doi.org/10.1128/genomeA.00187-14>.
7. Barrangou R, Fremaux C, Deveau H, Richards M, Boyaval P, Moineau S, Romero DA, Horvath P. 2007. CRISPR provides acquired resistance against viruses in prokaryotes. *Science* 315:1709–1712. <http://dx.doi.org/10.1126/science.1138140>.
8. Cai Y, Yan J, Hu X, Han B, Yuan Z. 2007. Improving the insecticidal activity against resistant *Culex quinquefasciatus* mosquitoes by expression of chitinase gene *chiAC* in *Bacillus sphaericus*. *Appl Environ Microbiol* 73:7744–7746. <http://dx.doi.org/10.1128/AEM.01510-07>.
9. Fedhila S, Nel P, Lereclus D. 2002. The *InhA2* metalloprotease of *Bacillus thuringiensis* strain 407 is required for pathogenicity in insects infected via the oral route. *J Bacteriol* 184:3296–3304. <http://dx.doi.org/10.1128/JB.184.12.3296-3304.2002>.
10. Nishiwaki H, Nakashima K, Ishida C, Kawamura T, Matsuda K. 2007. Cloning, functional characterization, and mode of action of a novel insecticidal pore-forming toxin, sphaericolysin, produced by *Bacillus sphaericus*. *Appl Environ Microbiol* 73:3404–3411. <http://dx.doi.org/10.1128/AEM.00021-07>.